LETTERS 2003 Vol. 5, No. 13 2291–2293

ORGANIC

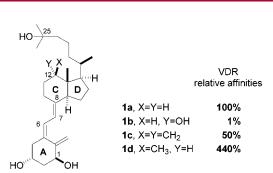
Functionalization at C-12 of 1α,25-Dihydroxyvitamin D₃ Strongly Modulates the Affinity for the Vitamin D Receptor (VDR)

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Received April 11, 2003



ABSTRACT

The first synthesis of analogues of the natural hormone $1\alpha_{2}$ -dihydroxyvitamin D_{3} (1a) with substituents at C-12 is reported. The following are the relative affinities of the novel compounds for the vitamin D receptor (VDR) compared to that of 1a (100%): $1\alpha_{1}$ - $12\alpha_{2}$ - $(OH)_{3}$ - D_{3} (1b, 1%), $1\alpha_{2}$ -2- $(OH)_{2}$ -12-methylene- D_{3} (1c, 50%), and $1\alpha_{2}$ -2- $(OH)_{2}$ - 12β -methyl- D_{3} (1d, 440%).

The steroid hormone 1α ,25-dihydroxyvitamin D₃ [1α ,25-(OH)₂-D₃, **1a**] is the bioactive metabolite of vitamin D₃. It plays an important role in the regulation of mineral metabolism and also promotes cell differentiation and inhibits the proliferation of various types of tumor cells, a fact that suggests its possible use in the treatment of cancer and other hyperproliferation diseases. Unfortunately, the therapeutic value of 1α ,25-(OH)₂-D₃ as an antitumor agent is severely limited by its potent calcemic activity.¹ Efforts aimed at developing vitamin D analogues with strong cell-differentiating ability and low calcemic action have led to the synthesis of more than 3000 analogues of 1α ,25-(OH)₂-D₃, and some of them show acceptable therapeutic profiles.² Most of these synthetic analogues have structural modifications at the side

chain² and A-ring,³ but only a few examples are known with modifications in the CD-rings⁴ and triene system.⁵

The hormone 1α ,25-(OH)₂-D₃ and the majority of its analogues exert their biological effects by binding with high affinity to the nuclear vitamin D receptor (VDR). Activation of the molecular switch of nuclear 1α ,25-(OH)₂-D₃ signaling, a process that starts with the formation of the complex composed of the ligand-activated vitamin D receptor (VDR),

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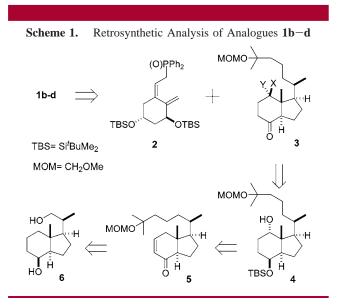
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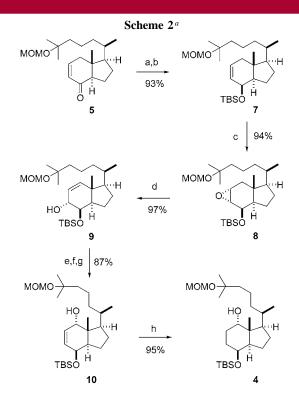
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the retinoid X receptor (RXR), and a 1α ,25-(OH)₂-D₃ response element (VDRE),⁶ leads to the regulation of the expression of various target genes, some of them involved in the regulation of cell growth, differentiation, and proliferation. Therefore, the ability of a vitamin D analogue (or mimic) to bind to the VDR is a prerequisite for the development of anticancer drugs that activate the vitamin D genomic mode of action. We recently embarked on a research program aimed at studying the structural requirements of the as yet unexplored C-12 position of the hormone 1α ,25-(OH)₂-D₃ for binding to the VDR. Here we describe the synthesis and VDR-binding properties of the first 1α ,25-(OH)₂-D₃ analogues bearing substituents at C-12, namely, 1α ,12 α ,25-(OH)₃-D₃ (**1b**), 1α ,25-(OH)₂-12-methylene-D₃ (**1c**), and 1α ,25-(OH)₂-12 β -methyl-D₃ (**1d**).⁷

The synthetic plan for the preparation of 1b-d involves the construction of the corresponding vitamin D triene units by the convergent Wittig-Horner approach (coupling between the anion of phosphine oxide 2 and ketones 3), and the preparation of the common intermediate 4 from Inhoffen-Lythgoe diol (6).⁸ This synthetic strategy offers a simple solution for the introduction of the 7,8-double bond of target vitamin D analogues but represents a challenge for the functionalization at C-12 (Scheme 1).



The α,β -unsaturated ketone **5** (Scheme 2) was prepared from Inhoffen–Lythgoe diol (**6**) as shown in earlier work.⁹ Stereoselective reduction of ketone **5** with diisobutylaluminum hydride and protection of the resulting alcohol with *tert*butyldimethylsilyl chloride provided **7**. Epoxidation of the double bond of **7** from the less hindered α face using *m*-chloroperbenzoic acid gave epoxide **8**. Attempts to open the epoxide ring with diethylaluminum-2,2,4,4-tetramethyl-



^{*a*} Reagents and conditions: (a) ^{*i*}Bu₂AlH, THF, -78 °C, 30 min (94%). (b) TBSCl, imidazole, DMF (99%). (c) *m*-CPBA, CH₂Cl₂ (94%). (d) LiNEt₂ (prepared from HNEt₂ and ^{*n*}BuLi, -78 °C to room temperature, Et₂O, 15 min), Et₂O, HMPA, rt, overnight (97%). (e) *m*-CPBA, CH₂Cl₂ (99%). (f) MsCl, Et₃N, CH₂Cl₂ (99%). (g) Na-naphthalene, THF, rt, 2 h (89%). (h) H₂, 5% Pd/C, EtOAc (95%).

piperidine¹⁰ failed. However, treatment of epoxide **8** with freshly prepared lithium diethylamide in the presence of hexamethylphosphoramide¹¹ provided the desired allylic alcohol **9**. Conversion of **9** into **10** was accomplished by a three-step sequence: (i) epoxidation with *m*-chloroperbenzoic acid, (ii) mesylation, and (iii) reaction with sodiumnaphthalene.¹² The stereochemistry of the resulting allylic alcohol **10** at C-12 was determined at the epoxyalcohol stage by NMR NOESY experiments. Catalytic hydrogenation of the double bond of **10** afforded the precursor **4** of the desired vitamin D analogues **1b**–**d** (70% overall yield from ketone **5**, eight steps).

With the alcohol **4** in hand, we decided to prepare the upper ketone fragments **3** required for the convergent synthesis of 1b-d. Conversion of **4** into **3a** (Scheme 3) began with desilylation ($^{n}Bu_{4}NF$, 89%), followed by selective oxidation of the C-8-OH group (PDC, 70%) and subsequent protective silylation of C-12-OH (TMSCl, 86%). The ketone **3b** was obtained in four steps from alcohol **4**. Oxidation of alcohol **4** (PDC, 95%), followed by olefination of the

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⁽⁷⁾ Steroidal numbering is used.

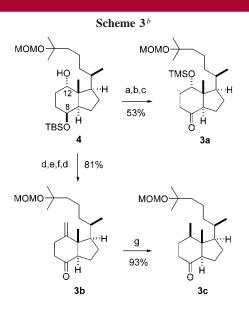
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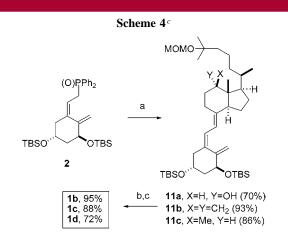


^b Reagents and conditions: (a) ${}^{n}Bu_{4}NF$, THF (89%). (b) Pyridinium dichromate (PDC), CH₂Cl₂ (70%). (c) Me₃SiCl, Et₃N, CH₂Cl₂ (86%). (d) PDC, CH₂Cl₂ (95%). (e) Ph₃PMeBr, KO'Bu, toluene (98%). (f) ${}^{n}Bu_{4}NF$, THF (92%). (g) H₂, 5% Pd/C, EtOAc (93%).

resulting C-12 ketone (Ph₃P=CH₂, 98%) and successive desilylation (*ⁿ*Bu₄NF, 92%) and oxidation (PDC, 95%), gave the C-8 ketone **3b** (81% yield from **4**). The ketone **3c** was prepared by catalytic hydrogenation of **3b** (5% Pd/C, 93%). NOESY studies established the C-12-Me of **3c** as having β stereochemistry.

Formation of the vitamin D triene units of protected analogues **11** (Scheme 4) proceeded under mild conditions by the coupling reaction between the anion of phosphine oxide **2** with the respective upper fragment **3**. Finally, deprotection of the hydroxyl groups of **11** ($^{n}Bu_{4}NF$, THF; AG50W-X4, MeOH) provided the desired 1α ,25-(OH)₂-D₃ analogues **1b**-**d**.

The receptor binding affinities of the novel vitamin D analogues **1b**-**d** were determined by in vitro competitive binding assays (RCI assay)¹³ using calf thymus vitamin D receptor. The following are the relative binding affinities of these compounds in comparison with that of the natural hormone 1α ,25-(OH)₂-D₃ (100%): **1b** (1%), **1c** (50%), and



 c Reagents and conditions: (a) $^n\text{BuLi},$ THF, 3, $-78\,^\circ\text{C.}$ (b) $^n\text{Bu}_4\text{NF},$ THF. (c) AG50W-X4, MeOH.

1d (440%). Noteworthy is the binding affinity of 1d, which is 4.4 times higher than that of the natural hormone 1α ,25-(OH)₂-D₃.

In summary, we have developed the first synthesis of C-12-substituted analogues of the natural hormone 1α ,25-(OH)₂-D₃. The presence of different substituents at this position dramatically changes the affinity of the resulting analogues for the vitamin D receptor. The high receptor binding affinity of analogue **1d** has encouraged us to develop new analogues in this series. The biological results and clinical potential of the new compounds will be reported in due course.

Acknowledgment. We are grateful to the DIGICYT (Spain, Project SAF 2001-3187) for financial support and to J.P. van de Velde and J. Zorgdrager (Solvay Pharmaceuticals BV, Weesp, The Netherlands) for the gift of starting materials and biological assays. X.C.G.-A. thanks the Spanish Ministry of Education and Technology for an FPI fellowship.

Supporting Information Available: Experimental procedures and spectral data (¹H and ¹³C NMR). This material is available free of charge via the Internet at http://pubs.acs.org.

OL034632C

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